

DIY PERSONAL CARE PRODUCT TESTING

Bacteria, Yeast & Mold

Quality Control

Check your cosmetic and personal care products for contamination using our Microslides®. It is fast, safe and easy!

Instructions

Carefully remove paddle from sterile vial and dip into a sample of a thin, aqueous solution, or apply creams and lotions to paddle with a sterile swab. Replace paddle in vial. Incubate or store at room temp.

Interpretation of Results

Check Microslide® after 24-48 hours of incubation or wait up to 5 days if stored at room temp. Includes evaluation chart for identifying level of contamination.



MICROBIAL TEST KIT

Microslides® contain a different test on each side of the paddle, for a 2-in-1 test. One side is selective for yeasts and moulds, while the other is selective for bacteria. This allows you to more easily identify contaminants.

- Microbiology training is not necessary
- For use on surfaces, liquids, & viscous materials, such as lotions or creams
- Economical price, simple application
- Available in kits of 4 or 10 Microslides®
- ✓ Shelf-life up to 9 months







Micro 4 pack: You will receive 10 sterile swabs

Micro 10 pack: You will receive 20 sterile swabs

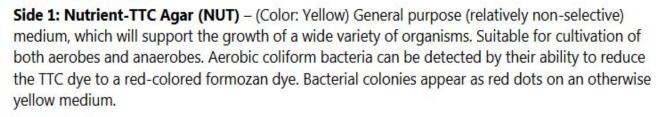
USE

Isolation and differentiation of Gram (-) enteric bacilli. (**NUT**) Selective enumeration and cultivation of yeasts, molds, and Actinomycetes from food and other surfaces (**RB**).

APPLICATION

In total coliform testing (TCC), the coliform organisms tested for include: total coliform, fecal coliform, and E. coli (Escherichia coli). Detection of fecal coliforms (a subset of total coliforms) or Escherichia coli (a subset of fecal coliforms) can indicate the potential presence of waterborne pathogens associated with fecal contamination¹. Rose Bengal Agar is recommended in *Standard Methods* for the enumeration of yeasts and molds from food and water.

PADDLE AGARS



Note: Paddle color is normally LIGHT YELLOW when the NUT agar is cast (about pH 6.0). Some microorganism growth (even before colonies are OBSERVABLE) will shift the pH from an acidic to a more alkaline level (pH 7.0 or higher) – turning the agar a light green.

Side 2: Rose Bengal Agar (RB) – (Color: Pink) Selective medium for the enumeration of yeasts and molds.

*Note: Side 1 of each paddle is marked with an indented laser line.



STORAGE / EXPIRATION

Microslides[®] should be stored tightly sealed (unopened) in a cool, dry location at room temperature (18 - 25°C; 65 - 77°F). Temperature fluctuations may result in condensation settling at the bottom of the vial, although this does not affect culture properties, it could reduce the shelf-life or cause the agar to separate from the plastic paddle support. Refer to 'Best Before End date' (SEE: BBE stamped on vial).

Avoid sudden temperature changes. Shield from direct sunlight. Do not allow paddles to freeze. Do not store in a refrigerator (~44°F / 10°C) or at temperatures exceeding 80°F; 27°C. Refrigeration may result in water condensation. Discard if paddle agar appears oxidized (darkened from expected color) or if contaminants appear. Expiry applies to medium in its intact container when stored as directed.

¹ United States Pharmacopeial Convention. 2007. The United States pharmacopeia, 31st ed., Amended Chapters 61, 62, 111. The United States Pharmacopeial Convention, Rockville, MD.



SAMPLING

SURFACE Sampling Protocol

- 1. Remove the paddle from the vial. Do not touch the agar surfaces.
- 2. To assure an accurate area recovery, contact the paddle to 20²cm of the surface by contacting the surface twice in separate 10²cm areas.
- 3. Replace paddle in vial.
- Incubate.

LIQUID Sampling Protocol

DIRECT IMMERSION PROTOCOL - low viscous liquids

- 1. Mix liquid test sample.
- 2. Remove the paddle from the vial. Do not touch the agar surfaces.
- When taking the sample:
 - a. Pour 40mL of the sample into the vial (to the printed horizontal fill line; see right). Dip the paddle into the 40mL volume liquid in the vial. Maintain a contact time of at least 15 seconds (30 seconds optimal). Both agar surfaces must be completely contacted.



- b. Or dip the paddle into the sample directly. Maintain a contact time of at least 15 seconds (30 seconds optimal). Both agar surfaces must be completely contacted.
- 4. Allow excess fluid to drain off both paddle agar surfaces.
- Replace paddle in vial.
- 6. Incubate.

SPREAD Protocol - high viscous liquids

- 1. Mix liquid test sample.
- 2. Remove paddle from vial. Do not touch the agar surfaces.
- 3. Holding the contact agar surface on a horizontal plane, deposit volume as a single drop approximately 1cm from the handle boundary (Figure 1).
- 4. Position a sterile glass rod on the "handle" side of the drop and bring it into contact with the drop creating a meniscus. Drag the glass tube over the paddle agar surface.
- Replace paddle in vial.
- Incubate.



Figure 1



INCUBATION

Incubation of Paddle Growth	Incubation Temperature	Examine at:
Yeast / Mold	25 to 30°C	48 hours up to 120 hours (5 days)
Yeast / Mold	Room Temperature	Up to 7 days
Total Coliform / Bacteria	35 ± 2°C	24 to 48 hours
Total Coliform / Bacteria	Room Temperature	Up to 5 days

Note: Incubation of bacteria after 48 hours may produce confluent growth making enumeration more difficult.

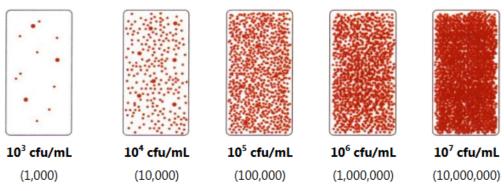
COLONY MEASURING

Each Microslide® paddle has molded media attachment points that are 4mm in length (point-to-point). This feature provides a useful guidepost to estimating nearby colony size.



ENUMERATION

Bacteria CFU/mL



Note: Estimation of lower counts is possible, but statistically difficult to justify. Use Light, Moderate and Heavy for Mold growth and surface testing.



DISPOSAL

Make a 1:9 dilution of household bleach (5.25% sodium hypochlorite solution). Twist and remove Microslide® paddle from vial. Fill vial with 40mL diluted hypochlorite solution (to fill-line). Allow 15-minute contact time. Discard bleach solution. Replace paddle in vial and dispose. Alternatively, loosen cap and microwave for 30 seconds, autoclave, or incinerate.

IDENTIFICATION

Organism	Nutrient-TTC (NUT)	Rose Bengal (RB)
Actinomyces bovis	Growth: + Colony: Opaque/tan-grey, CVEG, 1-3mm	Growth: ++ Colony: Opaque/tan-grey, CVEG, 1-3mm
Alternaria spp.	Growth: + Colony: Downy to wooly; flat, grayish, short, aerial hyphae, later becomes greenish black or olive-brown with a light border, 3-9cm	Growth: ++ Colony: Suede-like to woolly, initially white to yellow-orange, becoming black to olive- green or grayish, or grayish-green, umbonate with lighter center area, condication (rings), fast-growing, 3-9cm+ (confluent growth)
Aspergillus niger		
	Growth: +++ Colony: Granular, jet black conidia with yellow/gray hyphae, 3-5++cm	Growth: +++ Colony: Woolly and/or felt-like, forms a carpet, initially white later with jet black fruiting bodies (sporangia), fast-growing (4.5cm in 4 days), 3-9cm+ (confluent growth)
Aspergillus flavus	Growth: + Colony: Granular to wooly, yellow, yellow- green, or yellow-brown, 3-9cm	Growth: +++ Colony: Granular to wooly, yellow, yellow- green, or yellow-brown, 3-9cm+ (confluent growth)
Aspergillus fumigatus	Growth: + Colony: Granular to cottony, blue-green, green-grey, or green-brown, 3-9cm	Growth: +++ Colony: Felt-like, forms a carpet, initialy white to green or blue-green fruiting bodies, 3-9cm+ (confluent growth)
Aspergillus terreus	Growth: + Colony: Granular, radially rugose (wrinkled), cinnamon buff/brown, 3-9cm	Growth: +++ Colony: Granular, radially rugose (wrinkled), cinnamon buff/brown, 3-9cm+ (confluent growth)

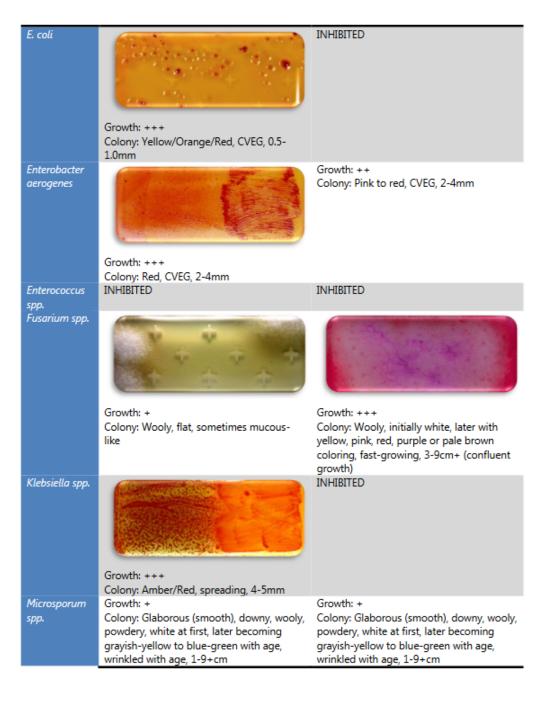




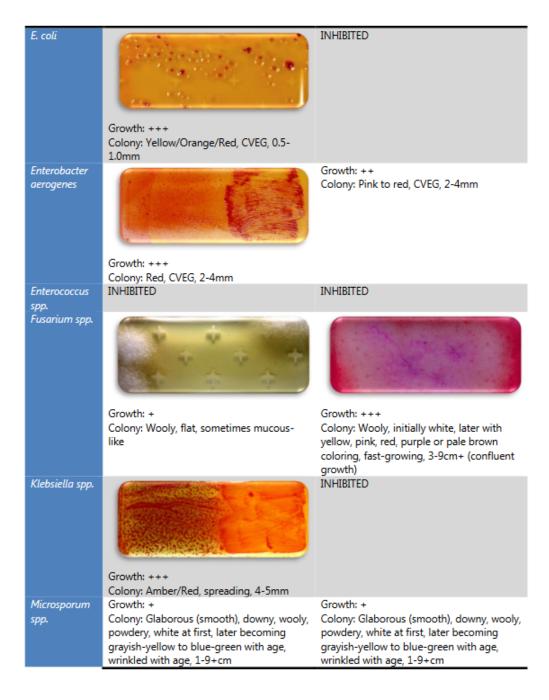


























	green or yellow-green patches (rings), 2- 9++cm	green or yellow-green patches (rings), 3- 9cm+ (confluent growth)
Trichophyton	Growth: +	Growth: ++
spp.	Colony: Wooly with indented boarders, white to brown/tan pigment, 2-9++cm	Colony: Wooly , initially white with brownish/tan pigmentation, outer darker ring, 3-9cm+
Gram (+) Bacteria	PARTIAL TO COMPLETE INHIBITION	

GLOSSARY

CVEG	Convex, Entire, Glossy
FED	Full, Entire, Dull
Gram	Gram reaction